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An alternative to Protein A for capture of Fc-containing bispecific antibodies

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Overview

- Introduction to CD3-TAA bispecific antibody Teneo-BsAb
- How to capture Teneo-BsAb?
- Initial test: Protein A
- Finding a better option: CaptureSelect CH1-XL
- Summary

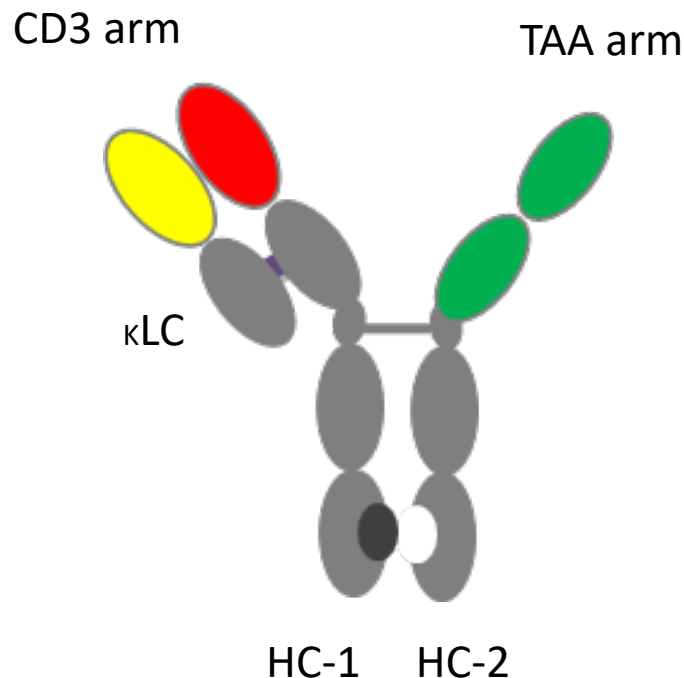


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Teneo-BsAb:

A CD3-tumor associated antigen (TAA)
bispecific antibody



TNB-BsAb CD3-TAA bispecific antibody

- Fully human IgG4 bispecific monoclonal antibody
 - 2 heavy chains (HC-1 and HC-2) and 1 kappa light chain (κLC)
 - Knobs into holes technology
 - Acid labile
- CD3 arm
 - HC-1 + κLC
 - T-cell receptor CD3
- TAA arm
 - HC-2 only
 - Consists of 2 identical VH domains recognizing TAA
 - Bivalent for increased avidity (<1 nM)
 - Derived from Teneobio's proprietary UniRat™ technology

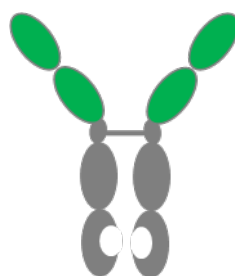
Teneo-BsAb CD3-TAA expressed products

Active



Heterodimer > 90%

Inactive

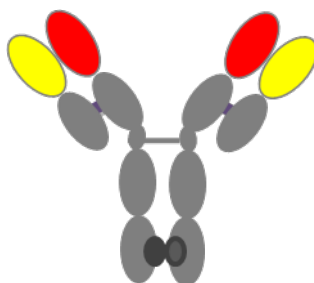


TAA Homodimer < 5%



Half-Ab < 1%

Aggregates < 2%

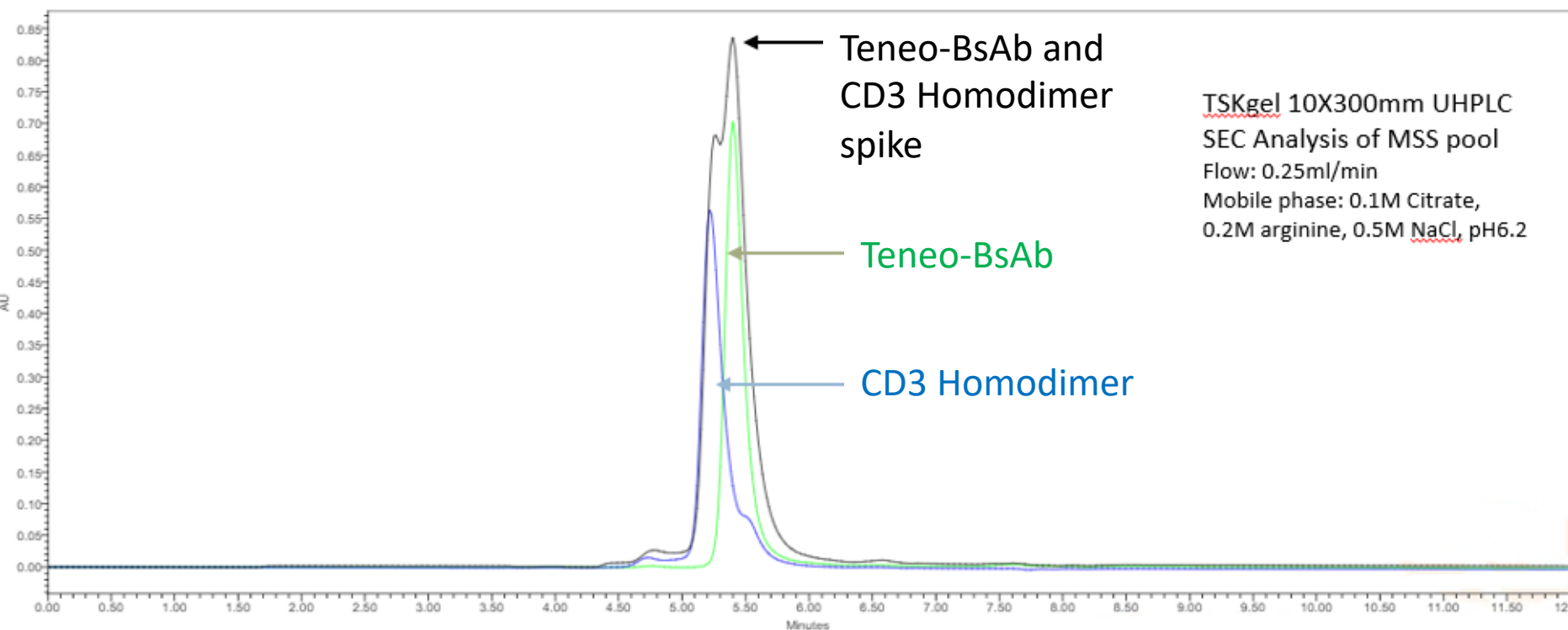


CD3 Homodimer < 5%

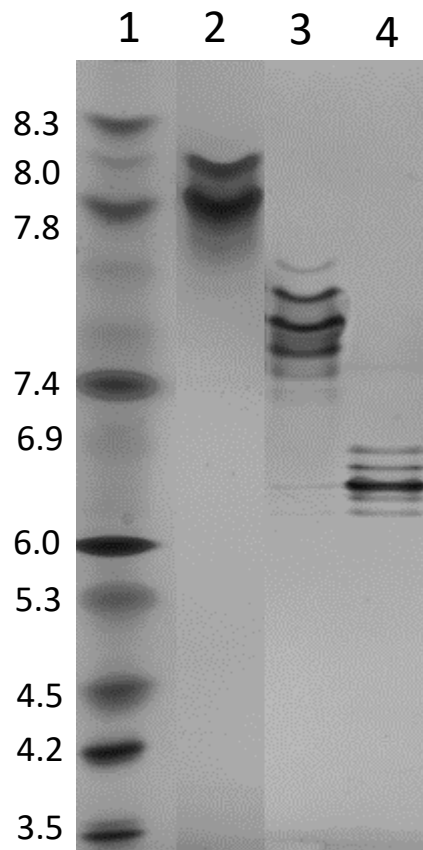


Excess LC

Teneo-BsAb heterodimer is similar in size to CD3 homodimers



Teneo-BsAb CD3-TAA heterodimer and homodimers have distinct pls



Lanes:

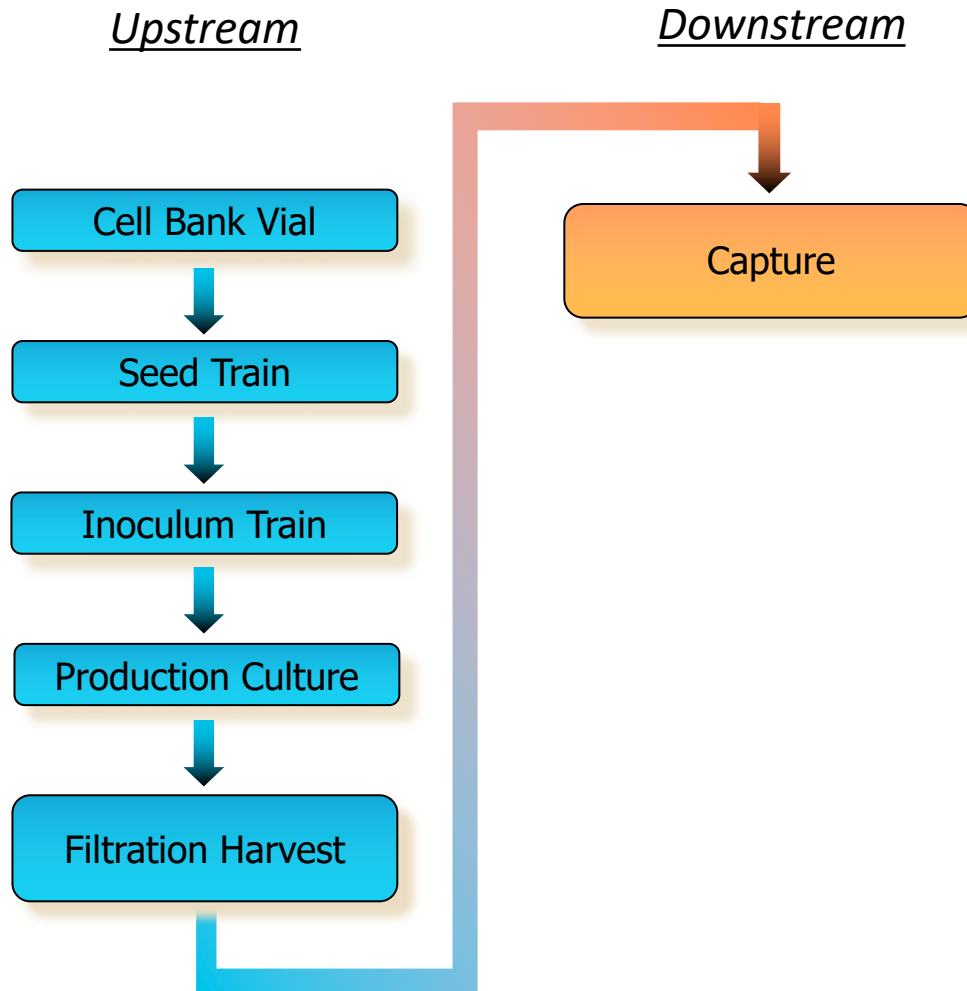
- 1) IEF, pI stds
- 2) CD3 Homodimer(knob-knob), pI = 8
- 3) Teneo-BsAb (Heterodimer) pI = 7.4-7.6
- 4) TAA Homodimer(hole-hole), pI = 6.2

5µg/lane loaded

pH 3-10 IEF gel(Invitrogen)
Instant Blue Stain(Expedeon)
Serva IEF Markers 3-10 mix
IEF Gel Program
1hr 100V 18mA 2.0W
1hr 200V 18mA 3.5W
30min 500V 18mA 9.0W

How to capture Teneo-BsAb?

Process for production of Teneo-BsAb CD3-TAA



Initial test for capture:

Protein A

Elution of Teneo-BsAb from Protein A at pH 3.6 is efficient...

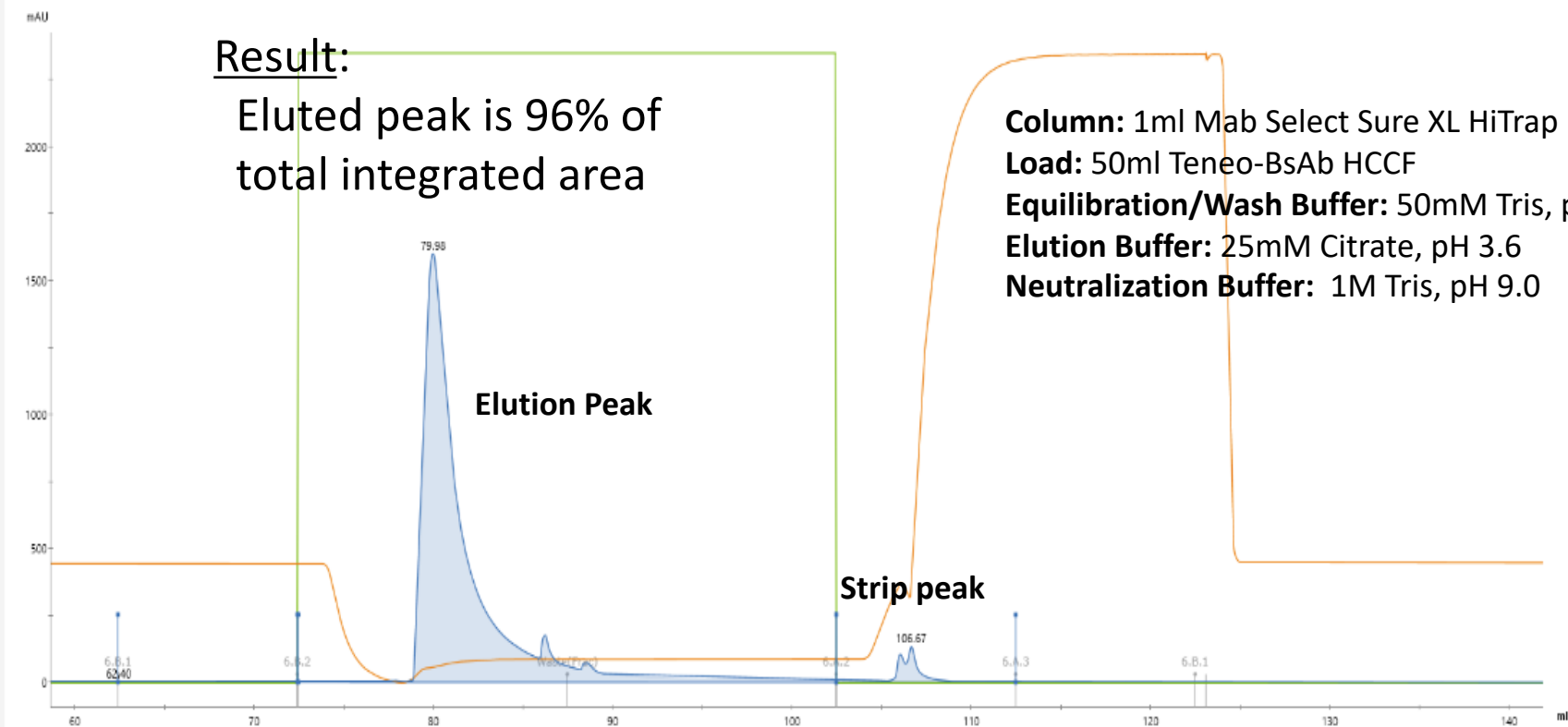
Result:

Eluted peak is 96% of total integrated area

Elution Peak

Column: 1ml Mab Select Sure XL HiTrap
Load: 50ml Teneo-BsAb HCCF
Equilibration/Wash Buffer: 50mM Tris, pH 7.0
Elution Buffer: 25mM Citrate, pH 3.6
Neutralization Buffer: 1M Tris, pH 9.0

Strip peak



Peak Table - UV

Peak	Retention ml	Area mAU	Area %	Ext coeff. mg ml ⁻¹ cm ⁻¹	Fraction(s)	Volume ml	Conductivity mS/cm
Peak A	62.404	5.214	0.12		6.B.1	10.002	4.47
Peak B	79.983	4093	96.02		6.B.3 - 6.A.1	30.000	1.17
Peak C	106.669	164.6	3.86		6.A.2	9.997	11.73

... But causes Teneo-BsAb to aggregate

Result:

SEC analysis indicates substantial amount of aggregated product after pH 3.6 elution

Column: Superdex200i 10/30 GL

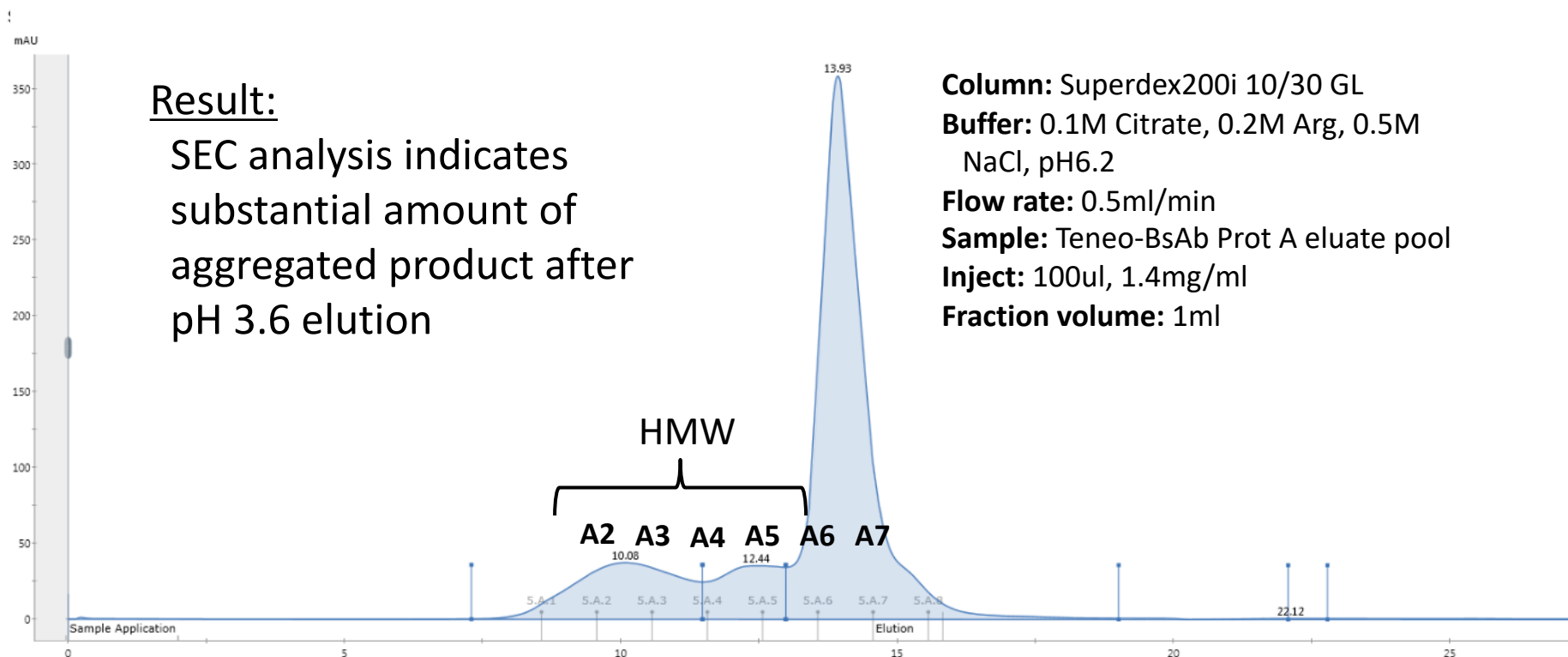
Buffer: 0.1M Citrate, 0.2M Arg, 0.5M NaCl, pH6.2

Flow rate: 0.5ml/min

Sample: Teneo-BsAb Prot A eluate pool

Inject: 100ul, 1.4mg/ml

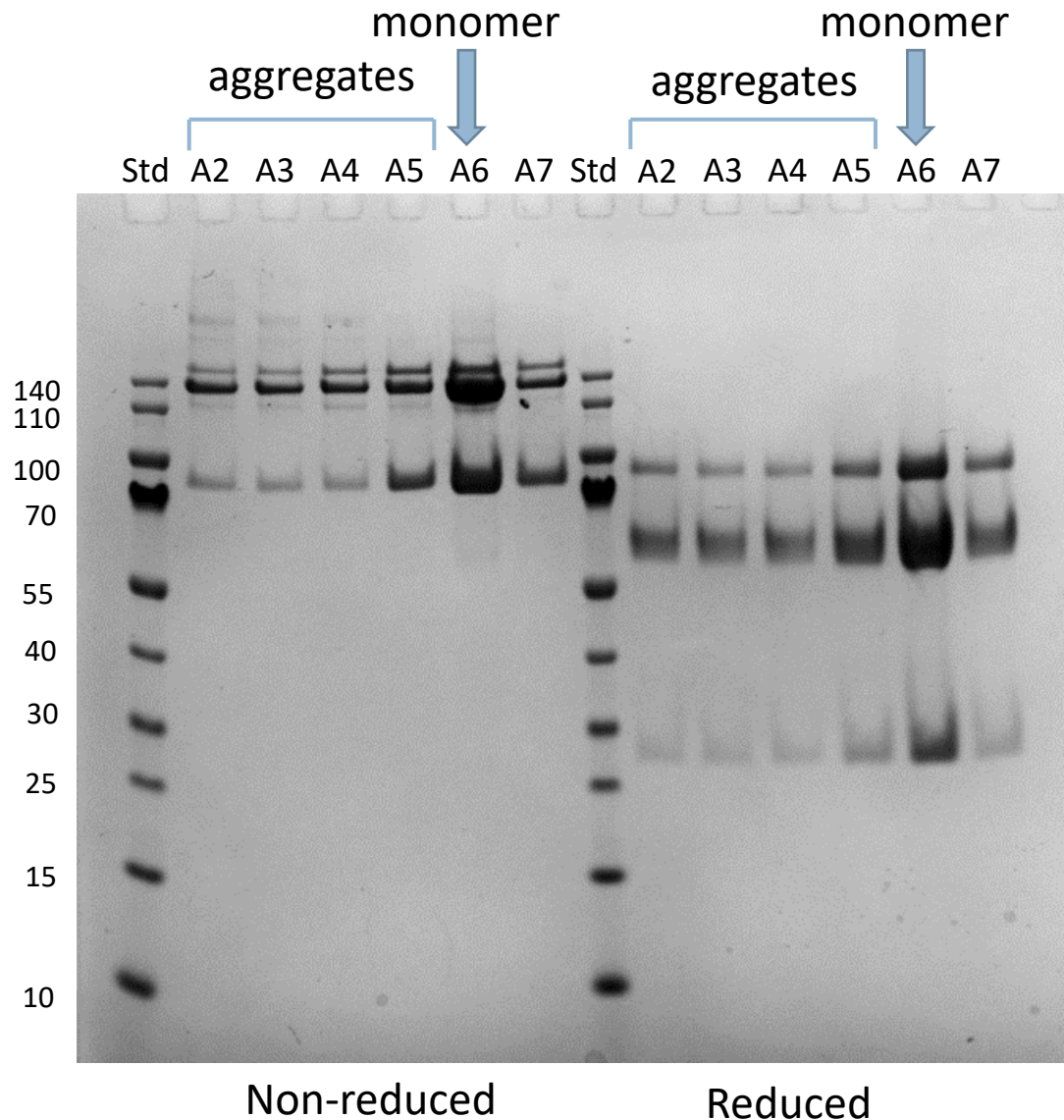
Fraction volume: 1ml



Peak Table - UV

Peak	Retention min	Area mAU	Area %	Ext. coeff. mg ml ⁻¹ cm ⁻¹	Fraction(s)	Volume ml	Conductivity mS/cm
Peak A	10.078	86.39	18.1		5.A.3	4.180	53.24
Peak B	12.444	48.28	10.11		5.A.3 - 5.A.5	1.508	53.23
Peak C	13.930	342.4	71.71		5.A.5 - Waste(Frac)	6.024	53.27
Peak D	22.118	0.3942	0.08		Waste(Frac)	0.713	53.20

SDS-PAGE confirms that HMW fractions are product



Protein A fractions

4-12% NuPAGE Gel
MES running buffer
5ug/lane load
Page Ruler Pre-----stain
Markers(Thermo)
Coommasie Stained

Can additives protect Teneo-BsAb at low pH?

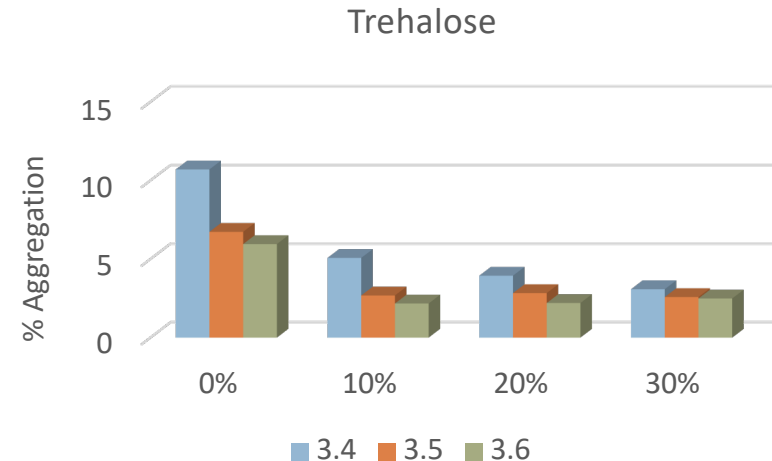
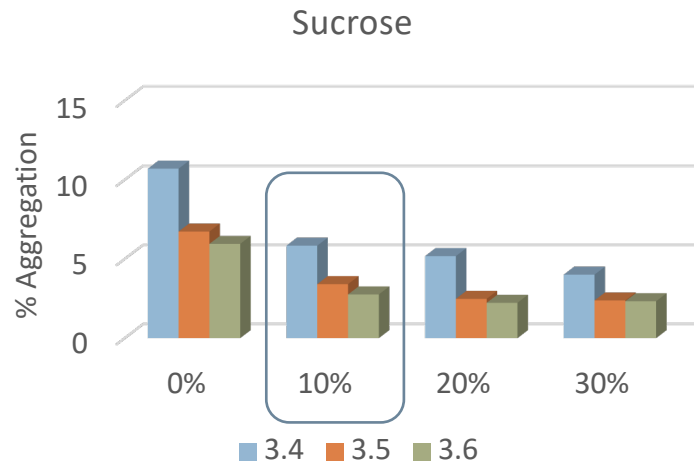
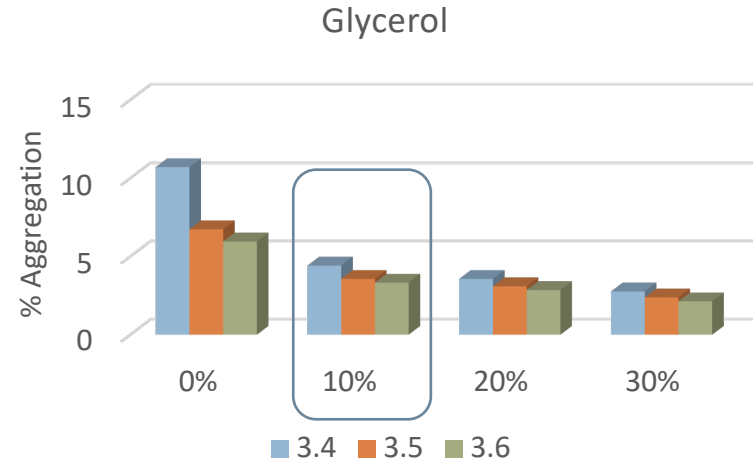
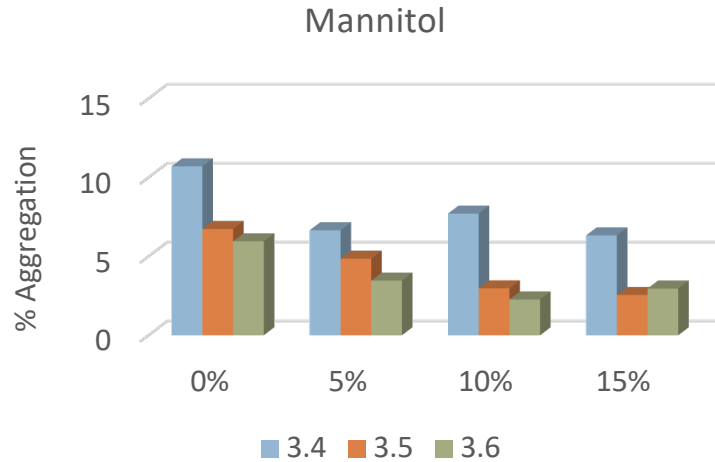
- DOE in GE Predictor Plates
- Elution buffer supplemented with polyols
- Test of three factors
 - Mannitol, glycerol, sucrose, trehalose
 - 5% - 30%
 - pH 3.4, 3.5, 3.6



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Additives can reduce aggregation of Protein A-eluted Teneo-BsAb CD3-TAA



Summary of issues with Protein A for capture

- Acid lability of Teneo-BsAb
 - Low pH elution will induce aggregation of product even with the addition of polyols to elution buffer
 - Cannot use low pH for virus inactivation
- Copurification of TAA - homodimer



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Finding a better option:

CaptureSelect CH1-XL

CaptureSelect CH1-XL

- General properties

- Ig heavy chain CH1-specific nanobody ligand
 - Eifler N., *Biotechnol. Prog.* **30**, 1311-1318, 2014
- Recognizes all four subclasses of IgG (IgG1, IgG2, IgG3, IgG4)
- Ligand immobilized on agarose 65 μ m
- Binding capacity < 20 mg/mL of IgG
- Flow 5 – 200 cm/hr
- Stable to base (25 – 50 mM NaOH) for sanitization
- Commercially available from Thermo

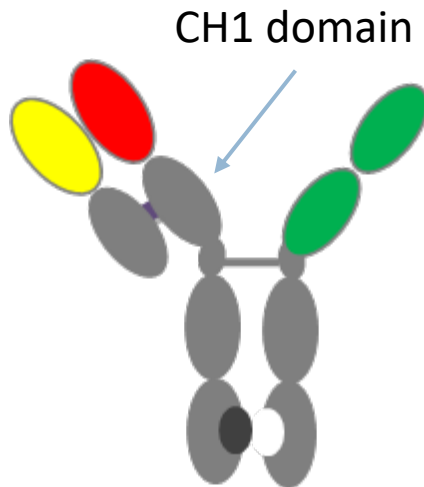
- For our specific need

- Binds bispecific heterodimer but NOT TAA homodimer
- Elutes under less stringent acidic (pH 4) condition



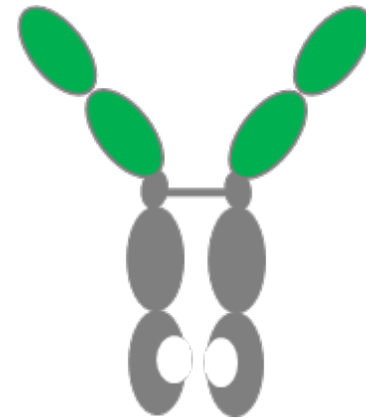
Only Teneo-BsAb contains a CH1 domain

Active



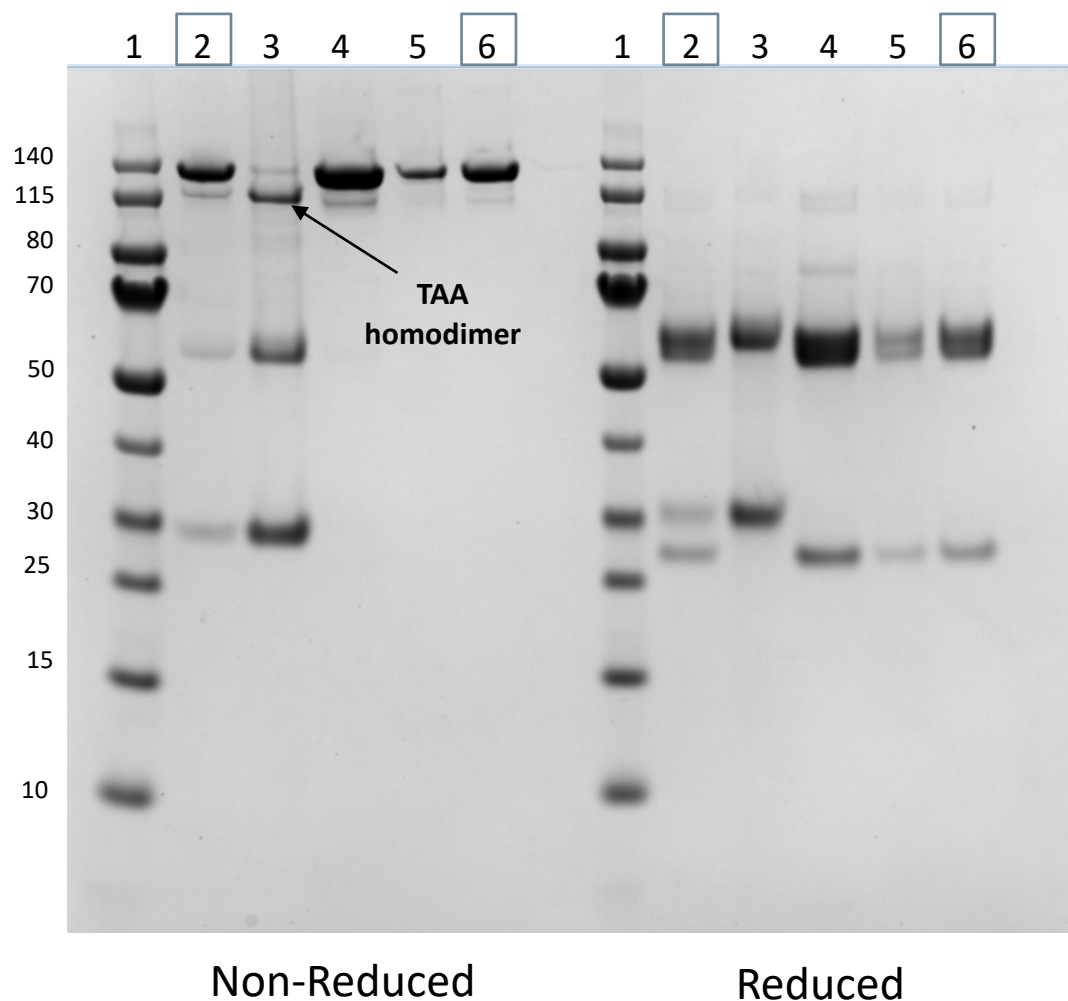
Heterodimer

Inactive



TAA Homodimer

Teneo-BsAb Protein A and CaptureSelect CH1 pools compared



Lanes:

- 1) MW stds:
- 2) Bispec.IgG ProA pool
- 3) Bispec.IgG CH1 **Flow trough**
- 4) CH1 salt wash:
- 5) CH1 NaOH strip:
- 6) CH1 pool:

Protein load: 2ug/lane

NuPAGE 4-12% Bis-Tris gel

MES Running Buffer

InstantBlue Stain (Expedeon)

PageRuler Prestained Protein Ladder

Run Conditions:

35min.

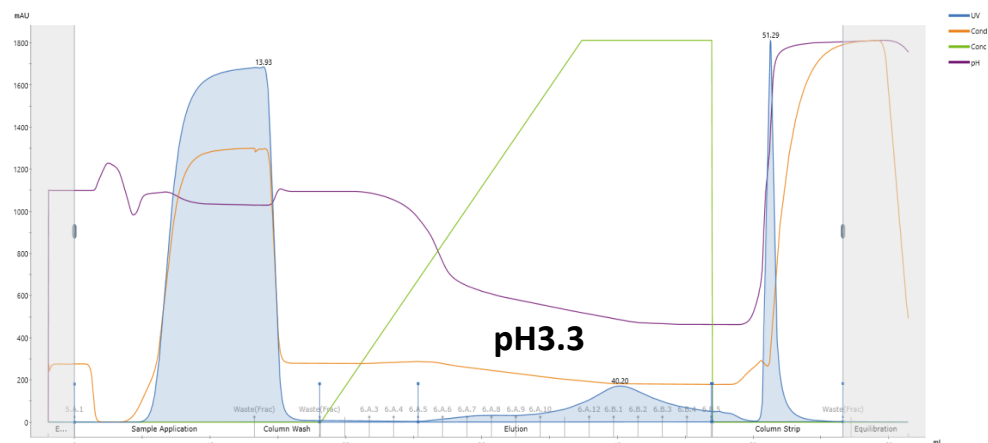
200V

120mA

25watts

Result: TAA homodimer is present in the CH1-XL flow through

Comparison of the elution pH of capture media



Column: 1ml Mab Select SuRe column

Buffers:

A- 50mM Tris, pH7.0

B-50mM Acetate, pH3.0

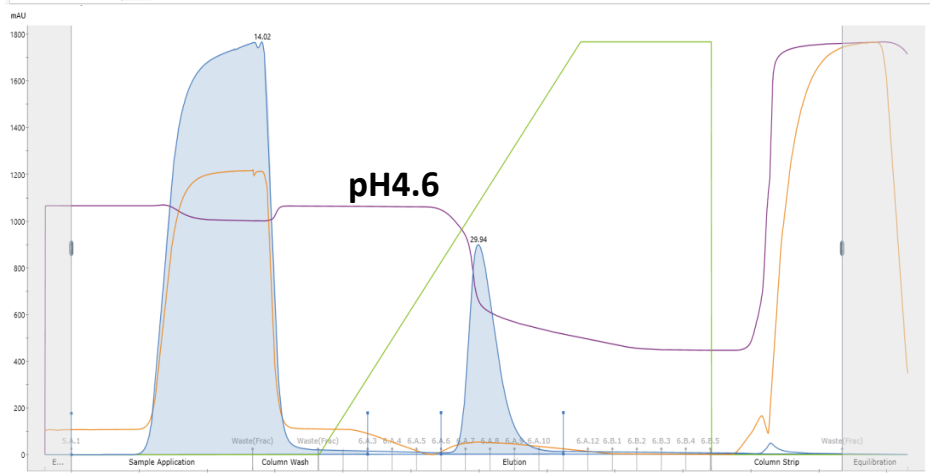
C- Strip Buffer: 0.1M NaOH

Elution: Linear grad. 10CV – 100%B

10ml HCCF load

Peak Table - UV

Peak	Retention	Area	Area %	Ext. coeff.	Fraction(s)	Volume	Conductivity
Peak A	13.829	12956	82.14	5.1-5.2	18.966	8.14	
Peak B	40.197	1439	9.19	6.4.5-6.4.6	21.001	2.72	
Peak C	51.288	1357	8.67	5.1.3	9.624	11.36	



Column: 1ml CaptureSelect CH1-XL

Buffers:

A- 50mM Tris, pH7.0

B-50mM Acetate, pH3.0

C- Strip Buffer: 0.1M NaOH

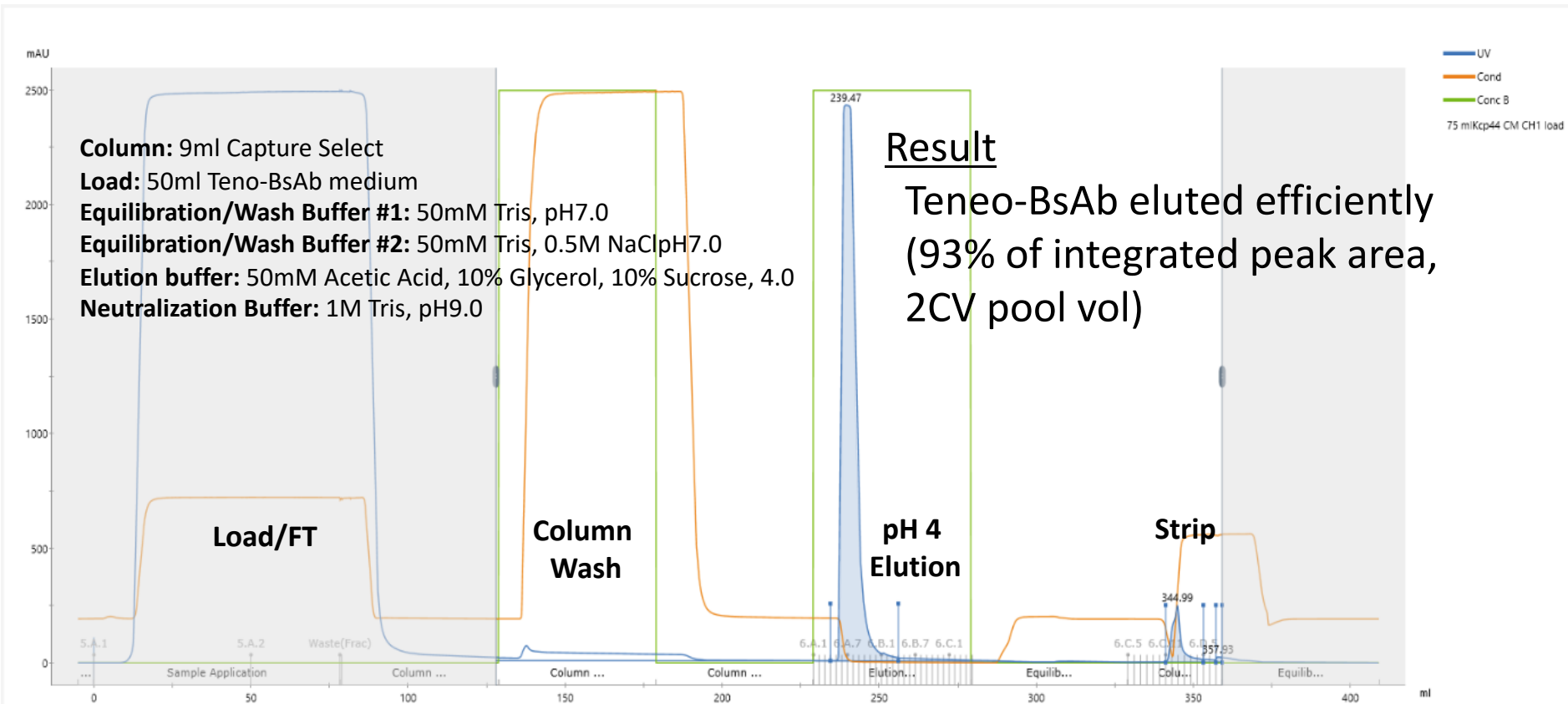
Elution: Linear grad. 10CV – 100%B

10ml HCCF load

Peak Table - UV

Peak	Retention	Area	Area %	Ext. coeff.	Fraction(s)	Volume	Conductivity
Peak A	14.016	13516	95.33	5.1-5.2	21.805	8.03	
Peak B	29.938	2140	13.67	6.4.6-6.4.10	9.000	2.75	

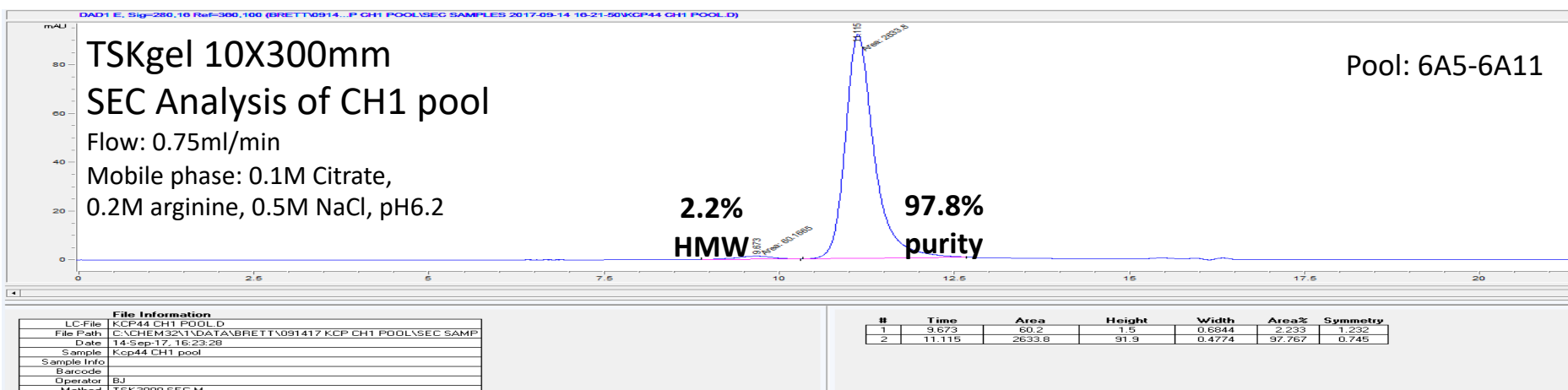
CaptureSelect CH1-XL elution of Teneo-BsAb at pH 4



Peak Table - UV

Peak	Retention ml	Area ml/mAU	Area %	Ext coeff. mg ml ⁻¹ cm ⁻¹	Fraction(s)	Volume ml	Conductivity mS/cm
Peak A	239.465	12113	93.21		6.A.4 - 6.B.3	21.602	1.31
Peak B	344.987	839.3	6.46		6.C.11 - 6.D.4	12.000	8.67
Peak C	357.934	43.83	0.34		6.D.7 - 6.D.8	2.009	11.86

Teneo-BsAb eluted from CH1 XL contains minimal aggregates



Result

CaptureSelect CH1-XL pool has low HMW content at 2.2% with efficient binding of product out of HCCF.

Preliminary dynamic binding capacity of CaptureSelect CH1-XL for Teneo-BsAb

Residence Time (min)	Dynamic Binding Capacity (mg/mL)
1	3.6
2	8.4
4	9.3
8	9.4

1ml CH1-XL column (0.7 X 2.5cm)
Load: Purified Teneo-BsAb 5mg/ml
Res. Times: 1,2,4,8 min
10% Breakout before elution
P.C.: by 280nm of pool

Result

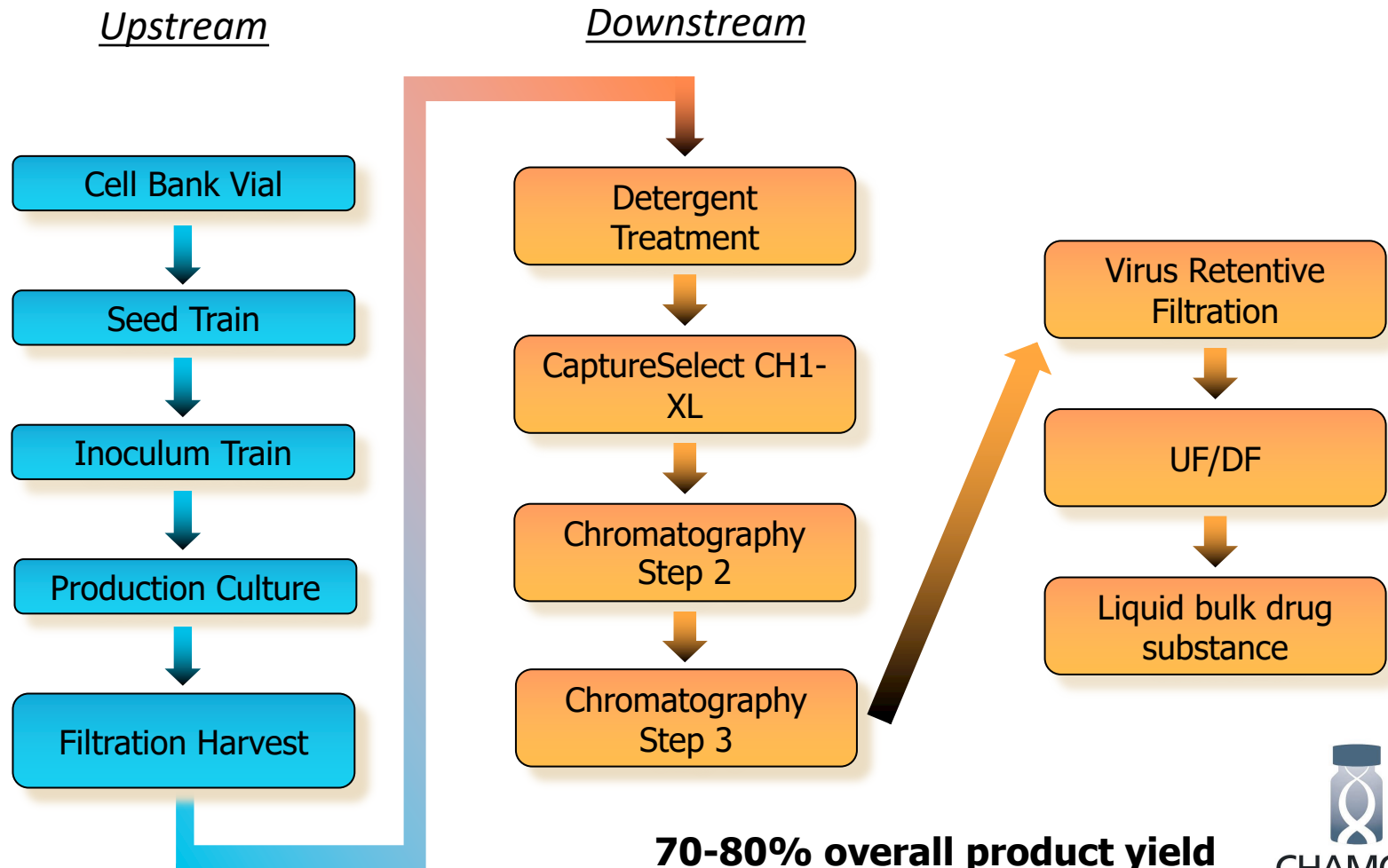
DBC plateaus at 4 min (9.3 mg/mL). Set minimum 4 minute residence time during loading of CaptureSelect CH1-XL.

NOTE:

Subsequent pilot-scale work using HCCF as load demonstrated the potential to increase load density (<19 mg/mL).

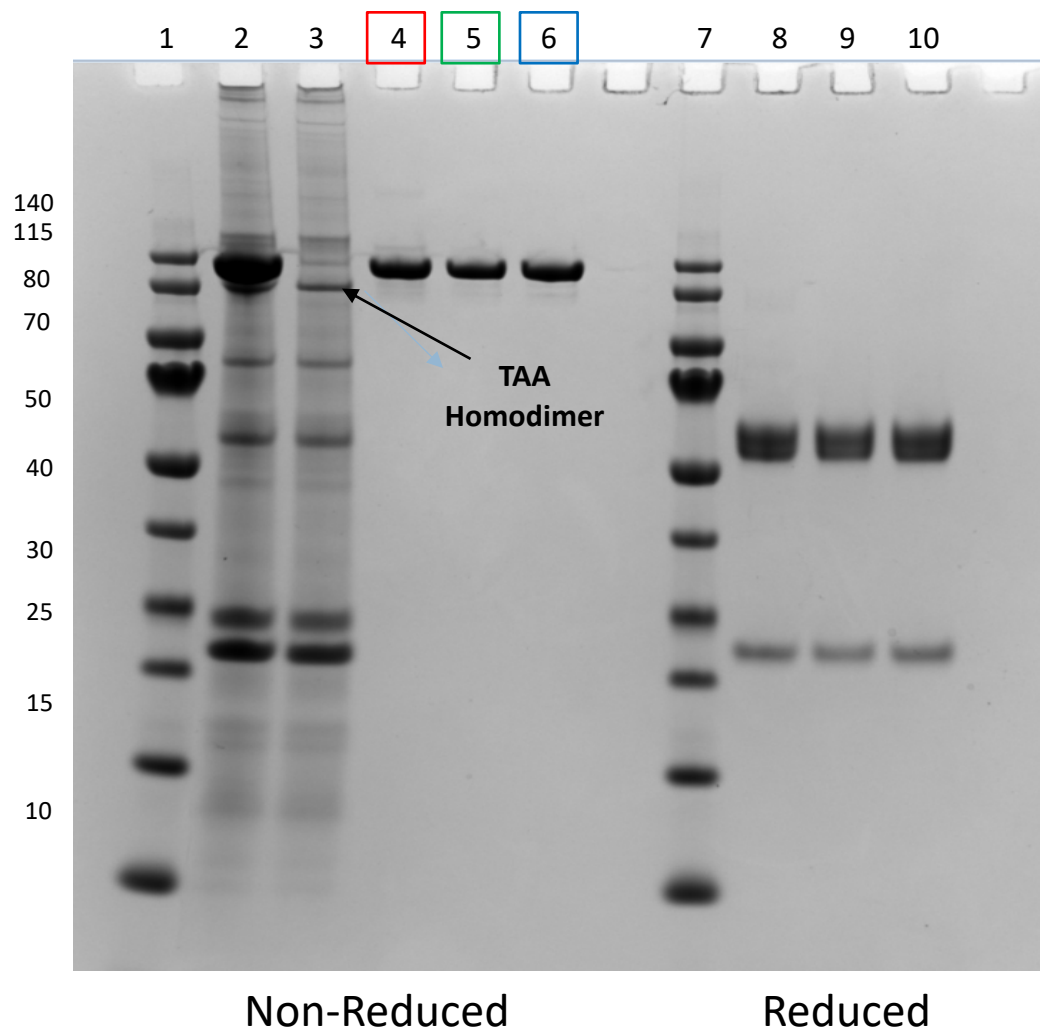


Process for clinical production of Teneo-BsAb drug substance



70-80% overall product yield

Summary of Teneo-BsAb purification through downstream process



Lanes:

- 1) MWS
- 2) HCCF 5ul
- 3) CH1 Flow through 5ul
- 4) CH1-XL1 pool 2ug
- 5) Purification step 2- pool 2 ug
- 6) Purification step 3- pool 2 ug
- 7) MWS
- 8) CH1 pool 2ug reduced
- 9) Purification step 2- pool 2 ug
- 10) Purification step 3- pool 2 ug

NuPAGE 4-12% Bis-Tris gel

MES Running Buffer

InstantBlue Stain(Expdeon)

PageRuler Prestained Protein Ladder

Protein load: 2ug/lane

Run Conditions:

35min.

200V

120mA

25watts



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Summary

- Teneo-BsAb is a CD3-TAA bispecific antibody that is trimeric, containing two H and one L chain
- We initially tested Protein A to capture
 - Low pH sensitivity of Teneo-BsAb was problematic
 - Copurification of TAA homodimer created additional downstream challenges



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Summary (cont'd.)

- CaptureSelect CH1-XL was identified as a better option
 - Mild elution condition (pH 4)
 - Good capacity (< 15 mg/mL)
 - High recovery (> 85% step yield)
 - However... low pH hold of eluted pool not an option for virus inactivation
- CaptureSelect could be more generally useful for bispecific constructs containing CH1 domains



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Thank you!

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